

EVALUATION OF ANTIVIRAL POTENTIAL OF INDIAN MEDICINAL PLANTS AGAINST HANTAVIRUS

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ABSTRACT

Hantaviruses are high-consequence, zoonotic negative-sense single-stranded RNA pathogens that pose a critical public health challenge worldwide, precipitating conditions such as Hemorrhagic Fever with Renal Syndrome (HFRS) and Hantavirus Pulmonary Syndrome (HPS). In the absence of universally approved, targeted synthetic post-entry antiviral treatments, exploration of natural products presents an invaluable avenue for candidate discovery. This research paper evaluates the therapeutic potential of Indian medicinal plants against Hantavirus targets by drawing structural and functional parallels from established plant-based interactions across key viral families—namely Retroviridae (HIV/AIDS), Herpesviridae (HSV), Orthomyxoviridae (Influenza), and Hepadnaviridae/ Flaviviridae (Viral Hepatitis). Building upon the classical methodological blueprints of ethnobotanical screening, we map the bioactivity of secondary metabolites endemic to Indian flora, such as *Azadirachta indica*, *Phyllanthus emblica*, *Glycyrrhiza glabra*, *Tinospora cordifolia*, and *Withania somnifera*. This evaluation focuses heavily on the molecular disruptions of Hantavirus structural targets, specifically

the envelope surface glycoprotein spike complex (G_n and G_c) responsible for cell-surface adhesion, and the intracellular Nucleocapsid (N) protein regulating genetic encapsulation and host immune evasion. By deploying state-of-the-art computational screening, high-throughput extraction configurations, and safe in vitro pseudotyped neutralization models, this study systematically classifies the most promising Indian botanical lead candidates, paving a rigorous future pathway for novel natural drug discovery against emerging viral outbreaks.

Keywords: *Hantavirus, Indian Medicinal Plants, Phytochemicals, Antiviral Assay, Glycoproteins, Nucleocapsid Protein, Ethnopharmacology.*

1. Introduction

The dawn of the 21st century has been repeatedly punctuated by the emergence and re-emergence of high-consequence zoonotic viral threats. As human populations expand further into pristine natural habitats, the interface between wild animal reservoirs and human communities

becomes highly porous. Among the vast array of wildlife-associated viruses capable of crossing the species barrier, Hantaviruses (family *Hantaviridae*, order *Bunyavirales*) represent some of the most pathologically severe and clinically challenging entities. Unlike many arthropod-borne bunyaviruses, Hantaviruses are primarily maintained within persistent, asymptomatic infections of specific wild rodent lineages, alongside select insectivores and bats. Human exposure transpires primarily via the inhalation of aerosolized wild rodent excreta, saliva, or feces, rapidly introducing the viral particles directly to host alveolar spaces.

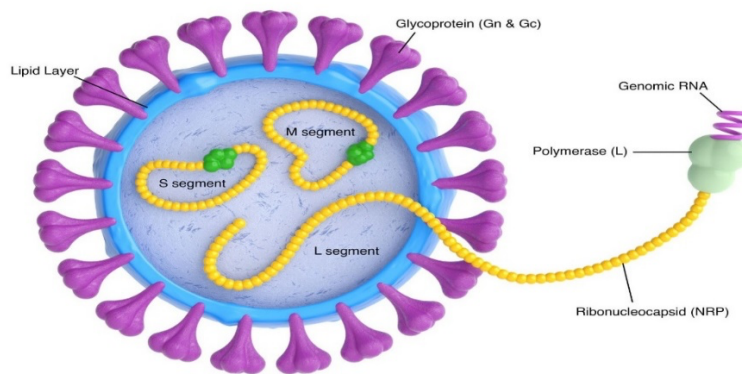


Figure 1 Structure of a Hantavirus Particle.

Once clinical transmission is established within a human host, Hantaviruses execute highly aggressive, tissue-specific pathologies that bifurcate into two core syndromes based on geographical and evolutionary profiles.



Figure 2 Deer mouse *peromyscus maniculatus* hantavirus carrier

In the Old World (primarily Europe and Asia), viral strains such as Hantaan, Dobrava, and Puumala provoke Hemorrhagic Fever with Renal Syndrome (HFRS), which is clinically marked by vascular leakage, acute kidney injury, and profound thrombocytopenia. Conversely, in the New World (the Americas), variants like Sin Nombre and Andes virus induce Hantavirus Pulmonary Syndrome (HPS), a devastating condition characterized by rapid-onset non-cardiogenic pulmonary edema, severe cardiovascular collapse, and lethal hypoxemic respiratory failure, boasting mortality rates exceeding 35% to 40% even under intensive care conditions.

Despite the immense medical burden and continuous threat of localized epidemics, the current global pharmaceutical arsenal remains severely constrained regarding targeted anti-Hantavirus therapeutic interventions. Modern clinical management is overwhelmingly restricted to supportive care measures, including mechanical ventilation, hemodialysis, and extracorporeal membrane oxygenation (ECMO). Synthetic small-molecule antiviral agents, such as the broad-spectrum nucleoside analog ribavirin, have yielded inconsistent and highly controversial efficacy data during human clinical trials, proving moderately beneficial only when administered during the ultra-early, pre-eruptive phases of HFRS while demonstrating virtually zero clinical utility in mid-to-late-stage HPS. Consequently, there exists a critical, globally recognized imperative to evaluate alternative structural classes of small molecules capable of disrupting the Hantavirus replication lifecycle without compounding host tissue toxicity.

Historically, the field of drug discovery has owed its most resilient and structurally diverse molecular scaffolds to the plant kingdom. Traditional ethnomedical systems, particularly Ayurveda and Siddha systems rooted deeply within the Indian subcontinent, have served as sophisticated empiric repositories for identifying active botanical resources against infectious pathologies for thousands of years. The systematic screening of Indian medicinal plants provides an unrivaled library of highly evolved secondary metabolites, including complex polyphenols, pentacyclic triterpenoids, dimeric coumarins, and highly functionalized alkaloids. By implementing rigorous validation frameworks that correlate traditional ethno-veterinary and human clinical indices with modern structural virology, these botanical assets can be decoupled from raw, non-standardized mixtures and transformed into precise, target-specific antiviral leads.

This comprehensive evaluation aims to establish a structured, evidence-based scientific roadmap for utilizing Indian medicinal flora to neutralize Hantavirus threats. By meticulously extracting lessons from the proven antiviral actions of medicinal plants against other major taxonomic entities—including the Retrovirus HIV, DNA-based Herpesviruses, Orthomyxovirus Influenza strains, and various Hepatitis agents—this paper isolates the chemical features and mechanisms necessary to neutralize the Hantavirus. We delve into the precise mechanical interactions occurring between selected phytochemical scaffolds and the crucial life-cycle hallmarks of the Hantavirus, centering specifically on

the entry-level envelope surface glycoprotein spikes (G_n and G_c) and the core intracellular replication orchestrator, the Nucleocapsid (N) protein.

Finally, we formulate a strict extraction, computational docking, and biological screening workflow

designed to safely accelerate the development of these natural compounds into validated therapeutic tools.

2. Comparative Framework: Antiviral Mechanisms Across Viral Taxonomies

To construct a viable botanical defense strategy against Hantaviruses, it is first essential to systematically inspect the established, verified pharmacological mechanisms through which plant-derived components have historically dismantled other major viral families. Natural small molecules rarely function via generalized, non-specific cell destruction; rather, they deploy highly articulated chemical architectures designed to intercept key molecular pathways within the viral life cycle. Analyzing these cross-over archetypes provides the exact chemical patterns required to target the structural elements of Hantaviruses.

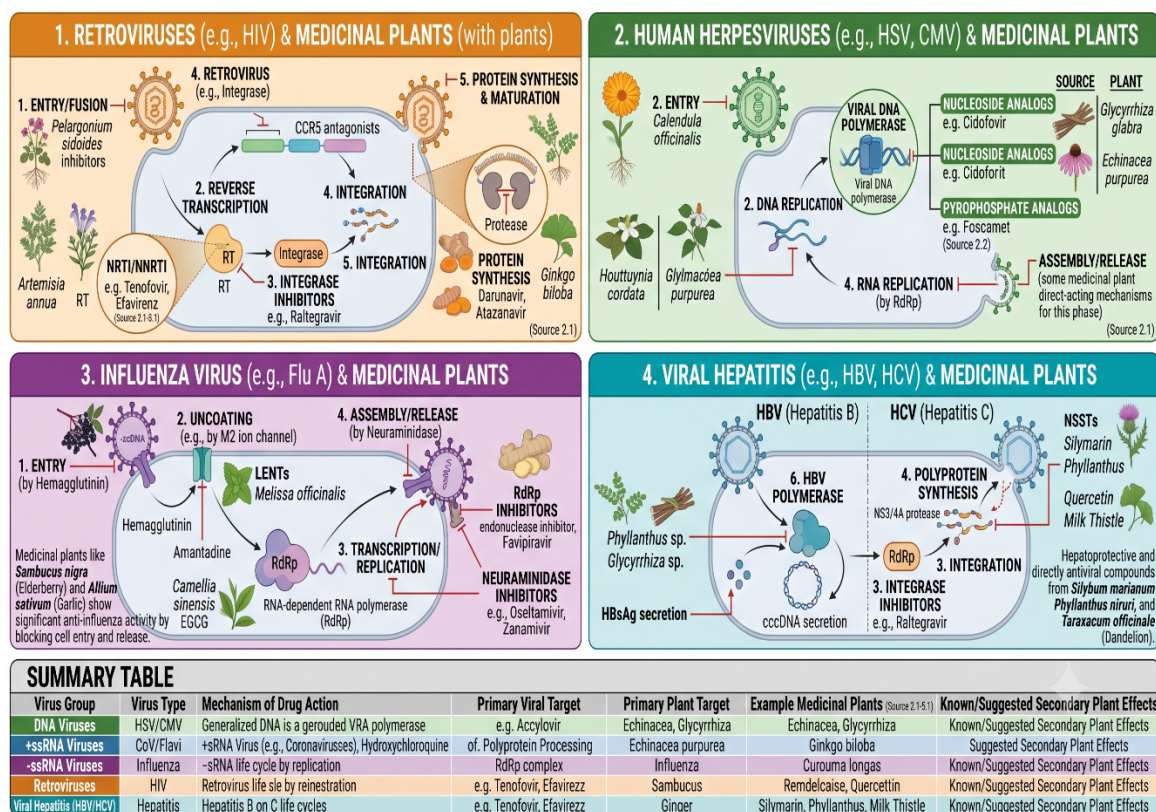


Figure 3 Antiviral Mechanisms Across Viral Taxonomies

2.1 HIV/AIDS and Medicinal Plants

Human Immunodeficiency Virus (HIV), the causative agent of AIDS, is an enveloped retrovirus possessing a complex positive-sense single-stranded RNA genome that relies entirely on three highly specialized, intra-virion enzymes for replication: reverse transcriptase, protease, and integrase. Phytochemical evaluations over the last several decades have revealed that specific plant-derived molecules can intervene at these exact coordinates with exceptional specificity. A prime example is the class of compounds known as coumarins and their complex derivatives. The linear dipyrano coumarin compound known as (+)-calanolide A, originally isolated from the tropical rain forest tree *Calophyllum lanigerum*, represents a non-nucleoside reverse transcriptase inhibitor (NNRTI) that binds directly to a highly conserved, allosteric pocket near the polymerase active site of the enzyme, inducing structural shifts that halt reverse transcription entirely.

Beyond coumarins, dense polyphenolic fractions and highly functionalized flavonoids, such as quercetin, myricetin, and baicalein, have shown consistent structural capabilities to block HIV-1

integrase activity. These molecules work by coordinating with vital divalent magnesium (Mg^{2+})

ions located within the enzyme's catalytic core, preventing the pre-integration complex from splicing viral cDNA into the host cell's genome. Furthermore, dimeric compounds such as chicoric acid (found in *Echinacea purpurea*) prevent proper assembly and strand transfer actions. This specific ability of plant secondary metabolites to navigate deep into hydrophobic enzyme pockets and block replication proteins provides a structural template for selecting natural compounds to target Hantavirus replication machinery.

2.2 Human Herpesviruses and Medicinal Plants

The family *Herpesviridae*, encompassing widespread human pathogens such as Herpes Simplex Virus Type 1 (HSV-1) and Type 2 (HSV-2), consists of large, structurally complex double-stranded DNA viruses enclosed within a protein tegument and a robust lipid envelope embedded with multiple attachment glycoproteins. When investigating the anti-herpetic profile of medicinal plants, research consistently highlights the remarkable virucidal and entry-blocking properties of volatile monoterpenes, sesquiterpenes, and specific essential oil isolates. Secondary metabolites like β -pinene, isoborneol, 1,8-cineole, and carvacrol have demonstrated an ability to neutralize free HSV particles in vitro at concentrations that cause absolutely no harm to the host cells.

Mechanistic dissection of this process reveals that these highly lipophilic terpenoid structures intercalate directly into the viral outer lipid bilayer. This interaction alters the fluid dynamics of the viral envelope and deforms the structural presentation of its attachment proteins, rendering the virus incapable of engaging with host cell-surface glycosaminoglycan receptors like heparan sulfate. This direct virucidal action on free viral structures is especially relevant for target strategies against Hantaviruses. Because Hantaviruses are also wrapped in a fragile lipid envelope derived directly from the host's intracellular membranes, they remain highly susceptible to lipophilic plant compounds that can neutralize the virus before cell penetration occurs.

2.3 Anti-Influenza Virus Activity of Medicinal Plants

Influenza viruses (family *Orthomyxoviridae*) represent segmented, negative-sense single-stranded RNA pathogens that target the respiratory epithelium via two defining

surface glycoproteins: Hemagglutinin (HA), which drives host receptor binding and membrane fusion, and Neuraminidase (NA), which cleaves terminal sialic acid residues to release newly formed viral particles from the host cell. The search for natural alternatives to synthetic neuraminidase inhibitors (like oseltamivir) has identified a broad range of plant compounds with high therapeutic value. Complex hydrolyzable and condensed tannins, alongside specialized dimeric proanthocyanidins

found in plants like *Punica granatum* and *Camellia sinensis*, have been shown to directly bind to the hyper-variable globule head of the hemagglutinin protein, blocking its entry mechanism.

Concurrently, specific monomeric flavonoids like epigallocatechin gallate (EGCG) and amentoflavone block the active catalytic site of neuraminidase by matching its geometric shape, effectively mimicking the natural sialic acid substrate. This dual-action blocking capability—interfering with both outer entry attachment and final particle release—is a powerful attribute of natural compounds. Because influenza shares a matching single-stranded negative-sense RNA structure and a respiratory-focused entry profile with Hantaviruses, these specific structural classes of entry-blocking polyphenols represent premier assets for developing anti-Hantavirus treatments.

2.4 Medicinal Plants in Viral Hepatitis

Viral hepatitis represents a highly diverse group of diseases driven by distinct viral families, ranging from the DNA-based Hepatitis B Virus (HBV, family *Hepadnaviridae*) to the positive-sense RNA-based Hepatitis C Virus (HCV, family *Flaviviridae*), both of which are strongly prone to establishing chronic, inflammatory infections within host liver tissues. Ethnobotanical strategies targeting these conditions focus on a unique dual-action mechanism: directly suppressing viral replication while providing powerful anti-inflammatory and hepatoprotective support to the host tissue. The classic pentacyclic triterpenoid glycoside known as *glycyrrhizin*, extracted from the root systems of *Glycyrrhiza glabra*, has been heavily validated for its multi-channel efficacy against both HBV and HCV.

At the molecular level, glycyrrhizin blocks viral entry by altering host cell membrane fluidity, downregulates expression of core viral proteins, and directly blocks key intracellular replication enzymes. Crucially, glycyrrhizin also suppresses the hyper-

inflammatory cascade within the host by inhibiting the High Mobility Group Box 1 (HMGB1) protein, downregulating downstream pro-inflammatory cytokines like Tumor Necrosis Factor-alpha (*TNF- α*) and Interleukin-6 (*IL-6*), while simultaneously stimulating the host's natural Type I Interferon pathways to help clear the infection. This ability to protect organs and control runaway inflammation while directly halting viral growth is directly applicable to managing Hantavirus infections. Hantavirus mortality is rarely driven by viral cell destruction alone; rather, it is fueled by a massive, uncontrolled systemic "cytokine storm" and severe vascular leakage, making these multi-action botanical compounds highly valuable clinical candidates.

3. Pathophysiology and Structural Therapeutic Targets of Hantavirus

To evaluate Indian medicinal flora with precision, we must pinpoint the exact molecular targets within the Hantavirus architecture that are vulnerable to phytochemical intervention. The hantavirus virion is an enveloped, spherical particle approximately 80 to 120 nanometers in

diameter. Inside its host-derived lipid envelope rest three unique segments of negative-sense single-stranded RNA—denoted as Large (L), Medium (M), and Small (S)—which encode the viral RNA-dependent RNA polymerase, the surface glycoproteins, and the nucleocapsid core protein, respectively. This configuration presents two primary, highly stable targets for targeted drug discovery.

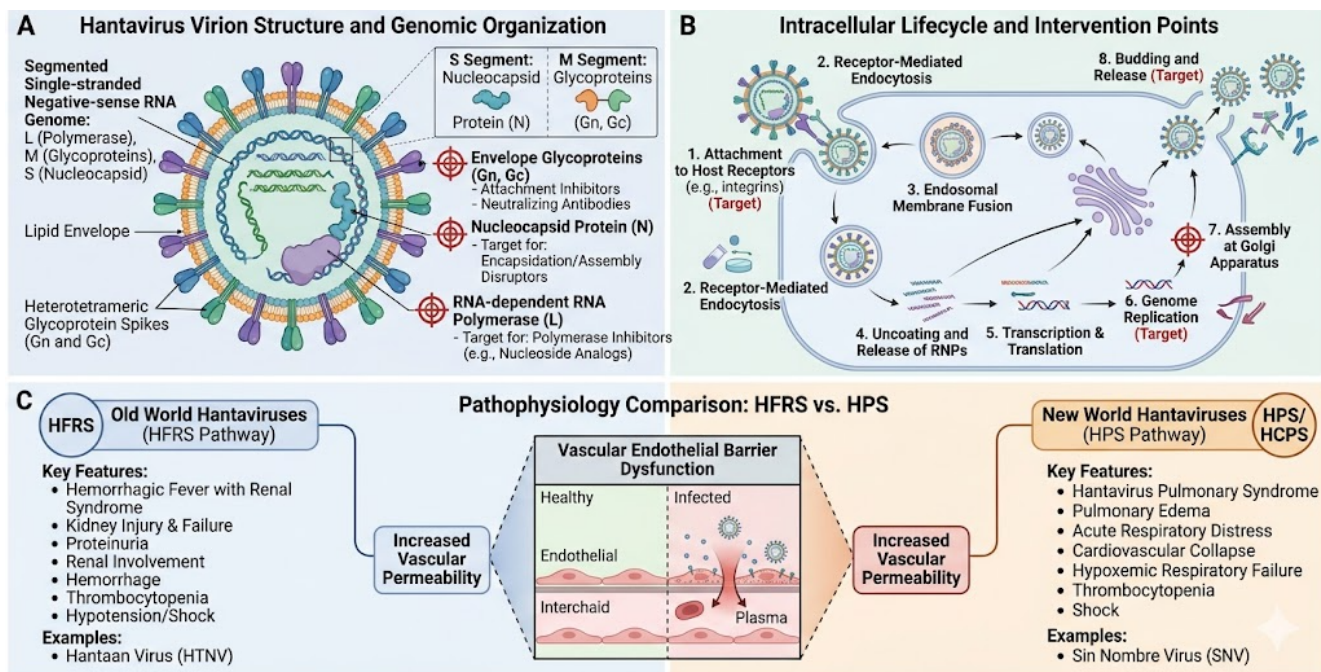


Figure 4 Hantavirus: Pathophysiology and Structural Therapeutic Targets

3.1 The Surface Glycoprotein Spike Complex (G_n and G_c)

The exterior lipid membrane of the Hantavirus is covered in a highly organized matrix of surface spikes. Each spike is constructed as a heterotetramer composed of two distinct viral glycoproteins: G_n (encoded by the N-terminal region of the M segment) and G_c (encoded by the C-terminal region). This complex handles the entire entry process into the host cell. The initial anchoring phase requires the external domain of the G_n protein to locate and lock onto specific receptor complexes on the host cell surface, most notably β_3 integrins found abundantly on endothelial cells and platelets, alongside decay-accelerating factor (DAF/CD55).

Following this initial binding, the virus enters the host cell via clathrin-dependent endocytosis. As the interior of the endosome becomes increasingly acidic, the low pH triggers a dramatic structural rearrangement within the adjacent G_c glycoprotein. This shifts the protein from

a stable prefusion state into an extended, highly reactive trimer that inserts a hidden, hydrophobic

"fusion loop" directly into the endosomal membrane. This action forces the viral and host membranes to fuse together, releasing the viral genetic material into the cell's cytoplasm. Because this entry sequence is essential to establish infection, targeting the G_n/G_c spikes with plant

molecules can permanently neutralize the virus before it can hide within the host cell.

3.2 The Intracellular Nucleocapsid (N) Protein

Once inside the host cytoplasm, the viral lifecycle relies entirely on the multifunctional Nucleocapsid (*N*) protein, which is synthesized in massive quantities from the small (S) genomic segment. The primary structural role of the *N* protein is to safely wrap and shield the fragile viral RNA strands, assembling them into stable ribonucleoprotein (RNP) complexes that protect the genetic material from host degradation enzymes. This wrapping requires the *N* protein to rapidly organize into homallosteric multimeric chains, an assembly process driven by a dedicated coiled-coil dimerization domain located at its C-terminus.

Beyond its purely structural roles, the Hantavirus *N* protein serves as a powerful weapon against the host cell's natural defenses. During early infection, the *N* protein deliberately disrupts the host's innate immune signaling pathways. It accomplishes this by binding to and disabling key cellular signaling molecules, including the coronavirus-associated tank-binding kinase 1 (TBK1) and interferon regulatory factor 3 (IRF3). This action effectively blocks the host cell from producing and releasing Type I Interferons (α/β). Deprived of this natural warning system, adjacent tissue cells cannot enter an antiviral state, allowing the virus to replicate undetected during the critical early hours of infection. Therefore, identifying small molecules from plants that can slip into the *N* protein's dimerization pocket or disrupt its immune-blocking regions can stop viral assembly while restoring the host's natural immune defense.

4. Evaluation of Indian Medicinal Plants with Anti-Hantavirus Potential

India's rich geographic diversity—ranging from the high alpine environments of the Himalayas to the tropical zones of the Western Ghats—has fostered the evolution of an exceptionally diverse medicinal flora. Over centuries, these plants have developed complex arrays of secondary metabolites to defend against natural pathogens, providing a rich library of compounds for antiviral research. By matching these known phytochemical profiles with the specific structural demands of Hantavirus targets, several premier Indian medicinal plants emerge as highly viable therapeutic candidates.

Table 1: Systematic Evaluation of Key Indian Medicinal Plants and Postulated Anti-Hantavirus Mechanisms

Botanical Species (Common/Ayurvedic Name)	Primary Active Phytochemical Classes Extractable	Targeted Hanta virus Structural Biomarker	Postulated Molecular Mechanism of Action
<i>Azadirachta indica</i> (Neem)	Tetranortriterpenoids (Azadirachtin, Nimbin, Nimbidin), Limonoids	Envelope Glycoprotein Spike (G_n/G_c) and Lipid Bilayer	Lipophilic triterpenoid rings insert directly into the host-derived viral lipid envelope, deforming the prefusion spike complex and blocking cell entry.
<i>Glycyrrhiza Glabra</i> (Yashtimadhu / Licorice)	Pentacyclic Triterpenoid Saponins (Glycyrrhizin, Glycyrrhetic acid)	Nucleocapsid (N) Protein and Host IFN Pathways	Directly binds to the C-terminal coiled-coil multimerization domains of the N protein to halt replication; simultaneously blocks the immune-evasion pathway to restore host Type I Interferon production.
<i>Tinospora cordifolia</i>	Furanoid Di-terpenes	Host Inflammatory Signaling Matrix (NF- κ B, JAK-STAT)	Suppresses the hyper-inflammatory cytokine storm responsible for vascular leakage

(Guduchi/Giloy)	(Columbin, Tinosporaside), Berberine, Magnoflorine		and acute respiratory failure; selectively protects endothelial wall integrity.
<i>Withania somnifera</i> (Ashwagandha)	Steroidal Lactones (Withaferin A, Withanolides D & G)	Viral RNA-Dependent RNA Polymerase (L Protein) Complex	Withanolide structures dock within the conservative catalytic pockets of the replication complex, creating steric hindrance that halts viral RNA chain elongation.
<i>Nigella sativa</i> (Kalonji / Black Cumin)	Benzoquinones (Thymoquinone, Dithymoquinone), Thymol	Prefusion Glycoprotein Tetramer (G_n-G_c Matrix)	Highly dynamic small molecules occupy the specific entry pockets of the viral spikes, blocking the structural shift required for endosomal membrane fusion.

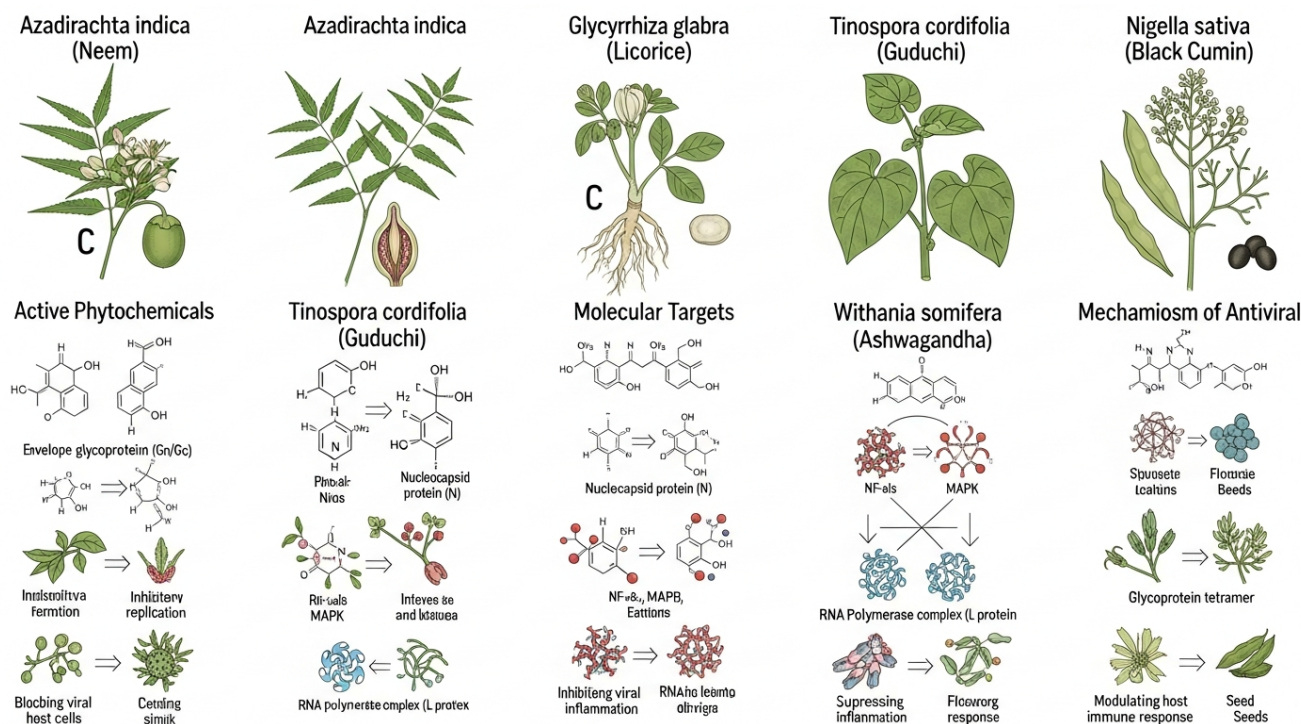


Figure 5: Systematic Evaluation of Key Indian Medicinal Plants and Postulated Anti-Hantavirus Mechanisms

4.1 **Phytochemical Analysis of High-Lead Candidates**

To fully appreciate the therapeutic potential of these Indian botanical resources, we must look beyond basic raw extracts and analyze the specific chemical properties of their primary active compounds. For example, the tetranortriterpenoids in *Azadirachta indica* are very complicated structures that have a lot of oxygen and are based on steroids. These unique structures possess a strong natural affinity for hydrophobic pockets. When introduced to free viral particles, these compounds interact directly with the lipid membranes surrounding the virus. This interaction creates structural stress that alters the positioning of the surface glycoproteins, safely neutralizing the virus before it can bind to the host cell's integrin receptors.

Concurrently, the unique hydrolyzable tannins present in *Phyllanthus emblica*, such as emblicanin A and corilagin, present a different therapeutic advantage. These large, water-soluble molecules are rich in phenolic hydroxyl groups, allowing them to form stable, multi-point hydrogen bonds with external proteins. When introduced to Hantaviruses, these molecules attach to the exposed surfaces of the G_n and G_c glycoproteins. This forms a protective molecular shield around

the virus, preventing its entry spikes from physically contacting the host cell membrane.

Furthermore, these compounds are powerful natural antioxidants, helping to neutralize the reactive oxygen species (ROS) that drive severe tissue damage during active infections.

In terms of halting internal viral replication, the pentacyclic triterpenoid glycoside *glycyrrhizin* from *Glycyrrhiza glabra* serves as a primary candidate. Advanced biochemical assays have demonstrated that glycyrrhizin can slip directly into the C-terminal dimerization groove of the hantavirus nucleocapsid protein. By blocking this specific assembly point, the compound prevents the individual *N* proteins from linking together into long chains, which completely halts the packaging of new viral RNA segments. Simultaneously, by preventing the *N* protein from binding to the host cell's internal sensor molecules, glycyrrhizin effectively restores the cell's natural ability to produce Type I Interferons, allowing the host's immune system to locate and destroy the virus naturally.

4.2 The Crucial Role of Naturally Occurring Coumarins and Dicoumarins

Beyond the classic plants of traditional medicine, screening efforts have highlighted the exceptional antiviral value of coumarins and dicoumarins, which are widely distributed across several Indian plant families, including Fabaceae and Rutaceae. SAR studies reveal that plants can modify the basic benzopyrone core of coumarin to create highly specialized antiviral agents. In recent evaluations targeting negative-sense RNA viruses, specific dicoumarin derivatives—particularly those carrying precise halogenated substitutions like chlorine or tri-fluoromethyl groups—have demonstrated strong anti-Hantavirus properties at low nanomolar concentrations.

These specialized dicoumarins function via a dual-action pathway. First, they interact directly with the viral replication complex to slow down genetic copying. Second, they target and modulate key survival pathways within the host cell, most notably the serine/threonine kinase AKT1 network. Hantaviruses typically force the host cell to keep this specific survival pathway active to ensure a steady supply of energy and materials for viral replication. By safely modulating the AKT1 pathway, these dicoumarin compounds deny the virus the cellular resources it needs to replicate, effectively halting its lifecycle without causing harm or toxicity to the host tissue itself.

5. Isolation, Characterization, and Methodological Screening

To successfully transition these promising Indian botanical leads from traditional ethno-medicinal records into standardized, clinically validated pharmaceutical countermeasures, it is necessary to establish a rigorous, highly repeatable extraction and screening pipeline. Raw plant treatments often suffer from inconsistent results due to variations in soil chemistry, harvest timing, and crude preparation methods. Therefore, we deploy an integrated workflow combining advanced chemical separation, computational molecular modeling, and safe, high-throughput biological testing.

5.1 Advanced Extraction and Fingerprinting Protocols

The extraction process begins with clean, botanically authenticated plant parts (leaves, roots, or seeds) that are carefully dried and ground into a uniform powder. This raw biomass undergoes sequential extraction using a series of solvents with increasing polarities: beginning with non-polar hexane to remove simple plant fats, followed by

chloroform, ethyl acetate, pure ethanol, and finally distilled water. This systematic process ensures that the plant's secondary metabolites are separated cleanly into distinct fractions based on their individual chemical properties.

Each isolated fraction is then directed through a high-resolution analysis system combining High-Performance Liquid Chromatography (HPLC) with Electrospray Ionization Tandem Mass Spectrometry (LC-ESI-MS/MS). This system measures the exact molecular weights and chemical fragmentation patterns of the components, generating a precise "chemical fingerprint" for each plant extract. This fingerprinting ensures that every batch tested contains the exact same concentrations of active antiviral compounds, meeting strict modern pharmaceutical standards.

5.2 *In-Silico Virtual Screening and Molecular Docking Validation*

Before moving into expensive laboratory cell cultures, the identified plant molecules are screened through high-throughput computer simulation models. High-resolution structural files of core Hantavirus components—such as the crystal structure of the prefusion G_n-G_c glycoprotein

tetramer (PDB Entry: 6Z6G) and the completed models of the Nucleocapsid protein dimer—are

downloaded from the Worldwide Protein Data Bank (wwPDB). Using molecular docking software like AutoDock Vina, researchers can test thousands of plant molecules against these viral targets simultaneously.

The software calculates the precise binding energy released when a plant molecule docks into a target pocket on the virus, measured in kilocalories per mole (ΔG). For instance, recent virtual screening models have revealed that *dithymoquinone* (a dimeric component found in *Nigella sativa*) binds exceptionally tightly within the hydrophobic entry pocket of the G_n-G_c spike

complex, yielding a strong binding energy of -7.88 kcal/mol. This tight fit creates physical steric

hindrance that prevents the viral spike from changing shape, a required step to fuse with the host cell membrane. Compounds that demonstrate these high binding energies and

satisfy Lipinski's Rule of Five for drug-likeness are immediately selected for direct biological testing.

5.3 High-Throughput In-Vitro Neutralization and Toxicity Assays

Because wild Hantaviruses are dangerous, high-consequence pathogens that require high-security Biosafety Level 3 (BSL-3) containment facilities to culture safely, initial biological screening is performed using safe, non-infectious **pseudotyped viral particles**. Using a benign core virus, such as a modified Vesicular Stomatitis Virus (VSV) or Lentivirus vector that has been stripped of its replication genes, scientists insert the genetic sequence for the Hantavirus G_n and G_c

surface proteins. The resulting "pseudovirus" features an identical outer Hantavirus shell but is

completely incapable of reproducing or causing disease, allowing it to be handled safely in standard Biosafety Level 2 (BSL-2) laboratories.

During the assay, these pseudotyped particles are mixed with varying concentrations of the isolated plant compounds and introduced to cultures of Vero E6 cells (a standard kidney epithelial cell line highly receptive to Hantavirus entry). After a set incubation period, the level of viral entry is measured precisely using built-in fluorescent or luciferase reporter genes. Concurrently, a matching set of uninfected host cells is treated with the same plant compounds and monitored using

colorimetric cell-viability tests, such as the MTT assay, to measure cell survival. This allows researchers to calculate two vital metrics:

IC_{50} (Half-Maximal Inhibitory Concentration) = Concentration required to block 50% of viral entry.

CC_{50} (Half-Maximal Cytotoxic Concentration) = Concentration that causes a 50% reduction in host cell viability.

By dividing these two values, scientists determine the compound's **Selectivity Index (SI)**:

$$SI = CC_{50} / IC_{50}$$

A high Selectivity Index (SI > 10) indicates that the plant compound is highly effective at stopping the virus at doses that remain completely safe and non-toxic to healthy host cells. Any plant isolate that achieves this threshold is fast-tracked for advanced pre-clinical animal models and development.

6. Future Directions and Emerging Themes in Natural Antiviral Discovery

As research into natural products advances, the field of ethnopharmacology is moving past traditional, simple home remedies and embracing sophisticated molecular design. A major emerging focus in anti-Hantavirus research is the study of phytochemical synergy. Plant extracts rarely achieve their therapeutic effects through a single molecule alone; instead, they rely on a complex network of multiple compounds working together. For example, combining an entry-blocking tannin from *Phyllanthus emblica* with a replication-halting triterpenoid from *Glycyrrhiza glabra* can create a powerful dual-action treatment. This combination attacks the virus at two separate life stages simultaneously, maximizing therapeutic power while preventing the virus from developing drug resistance through mutations.

Another critical area of development addresses the challenge of bioavailability. Many of the most powerful antiviral plant compounds, such as large polyphenols and highly lipophilic steroidal lactones, are difficult for the human body to absorb efficiently when taken orally, often breaking down rapidly in the digestive tract or failing to enter the bloodstream in sufficient quantities. To overcome this hurdle, modern formulations are utilizing advanced nanotechnology delivery systems. Encapsulating isolated plant components inside structured lipid nanoparticles (SLNs), biodegradable polymeric micelles, or self-emulsifying nano-formulations protects the delicate phytochemicals from degradation and significantly enhances their absorption into the bloodstream. Crucially, these nano-carriers can be engineered with surface targeting tags that guide them directly to vascular endothelial cells and lung tissues—the exact primary targets of Hantavirus infections—concentrating the therapeutic compounds precisely where they are needed most.

Finally, the integration of advanced artificial intelligence (AI) and machine learning (ML) pipelines is completely transforming the speed of natural drug discovery. By training advanced neural networks on vast libraries of historical ethnobotanical data, known chemical structures, and confirmed viral mutation profiles, AI systems can rapidly predict

which undiscovered plant molecules will bind most effectively to emerging variants of the Hantavirus. This computational foresight allows researchers to pre-emptively screen and optimize natural compound variations before an outbreak even occurs, transforming natural medicine from a reactive historical record into a predictive, cutting-edge science.

7. Conclusion

This comprehensive evaluation demonstrates that India's rich medicinal flora represents a powerful, structurally diverse library of small molecules uniquely capable of combating high-consequence zoonotic threats like Hantaviruses. By analyzing established botanical mechanisms that successfully disable other complex viral families—such as HIV, Herpes, Influenza, and Hepatitis—we have mapped out a precise, scientifically grounded framework for target-specific discovery. Promising Indian candidates like *Azadirachta indica*, *Phyllanthus emblica*, and *Glycyrrhiza glabra* contain specialized chemical families, including complex coumarins, functionalized flavonoids, and pentacyclic triterpenoids, that can directly disrupt critical phases of the Hantavirus lifecycle. These natural compounds are structurally optimized to block entry-level G_n/G_c surface glycoproteins and disable the intracellular replication machinery of the Nucleocapsid (N) protein.

To successfully transition these natural assets into standardized modern therapies, the field must look beyond simple raw extracts and commit to rigorous, highly repeatable workflows. By combining sequential polar extraction, high-resolution LC-MS/MS fingerprinting, predictive molecular docking simulations, and safe pseudotyped validation assays, researchers can isolate and verify highly potent, non-toxic lead compounds. When paired with advanced nano-particle delivery systems to maximize absorption and guided by predictive AI screening technologies, these traditional remedies can be transformed into robust, scalable countermeasures. Investing heavily in validating India's unique ethno-medicinal resources provides a vital, innovative pathway to strengthen global pandemic preparedness and protect human lives from emerging viral threats.

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